# Caesium uptake by roots of Medicago plants



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## Summary

- Medicago plants were grown with 1.65 and 20 mM
- Caesium inhibits growth of low potassium and high potassium plants
- Low potassium plants are more sensitive to caesium
- Caesium competes with potassium for uptake

### Introduction

- Potassium (K) is an essential element for plant growth and development
- Caesium (Cs) is chemically similar to K, but Cs is toxic to
- Because of the chemical similarity between Cs and K, root uptake mechanisms cannot differentiate between these elements easily (White & Broadley 2000)



### Materials & Methods

An in vitro system was used to grow Medicago truncatula (Figure 1). The plants were grown on MSR medium (Declerck et al. 2003) containing either 1.65 mM K (low K) or 20 mM K (high K). Various



Figure 1: In vitro system to grow Medicago truncatula et al. 2006).

(Dupré de Boulois

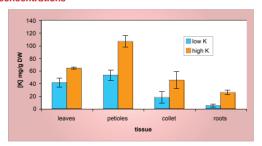
concentrations of CsCl

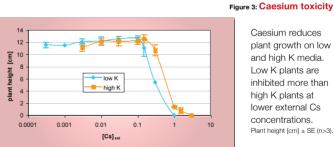
were added to these media. After 6 weeks the plants were harvested, oven dried and acid digested in a microwave. Concentrations of elements were measured using ICP-MS (PerkinElmerSCIEX, Massachusetts, USA).

### Results

### Figure 2: Potassium concentrations

Plants grown on low K and high K media differ in tissue K concentrations Mean K concentrations [mg/g DW] ± SE (n>3).



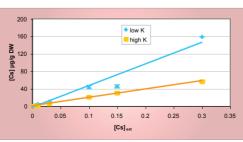


Caesium reduces plant growth on low and high K media. Low K plants are inhibited more than

high K plants at lower external Cs concentrations Plant height [cm] ± SE (n>3).

#### Figure 4: Caesium uptake

Plants take up Cs. Low K plants accumulate more Cs than high K plants at the same external Cs concentration. Shoot Cs concentrations [µg/g DW] ± SE (n>3).



### Figure 5: Competition for uptake

150 Ξ [Cs]<sub>ext</sub>

Caesium competes with K for uptake. High K plants have higher K concentrations in their tissues when grown at the same external Cs concentration than low K plants Shoot K concentrations [mg/g DW] ± SE (n>3).

Declerck S et al., 2003. Environmental Microbiology 5, 510-516 Dupré de Boulois H et al., 2006. Environmental Microbiology 8, 1926-1934 White PJ & Broadley MR, 2000. New Phytologist 147, 241-256

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